

# Hierarchical control of swelling in pulp fibers

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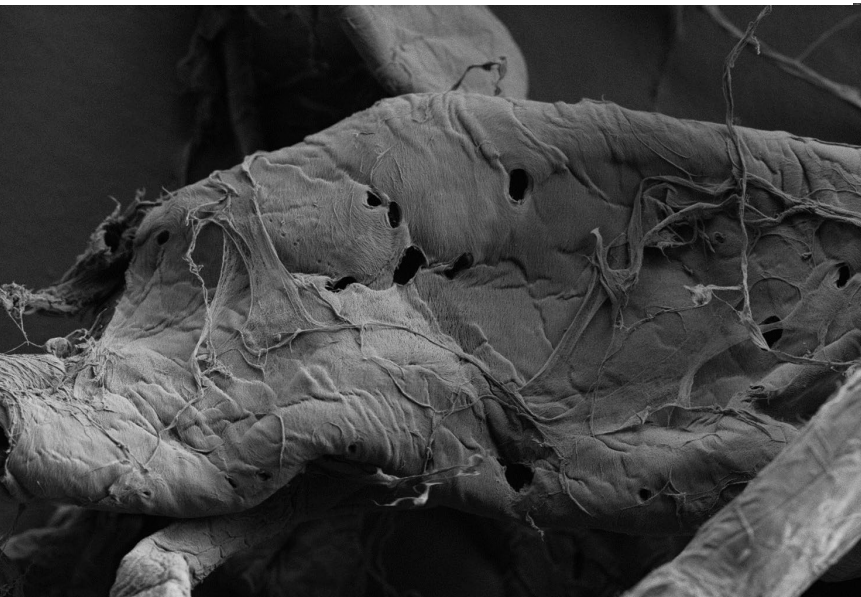


# Introduction

Fiber/water interactions → fiber swelling → product performance

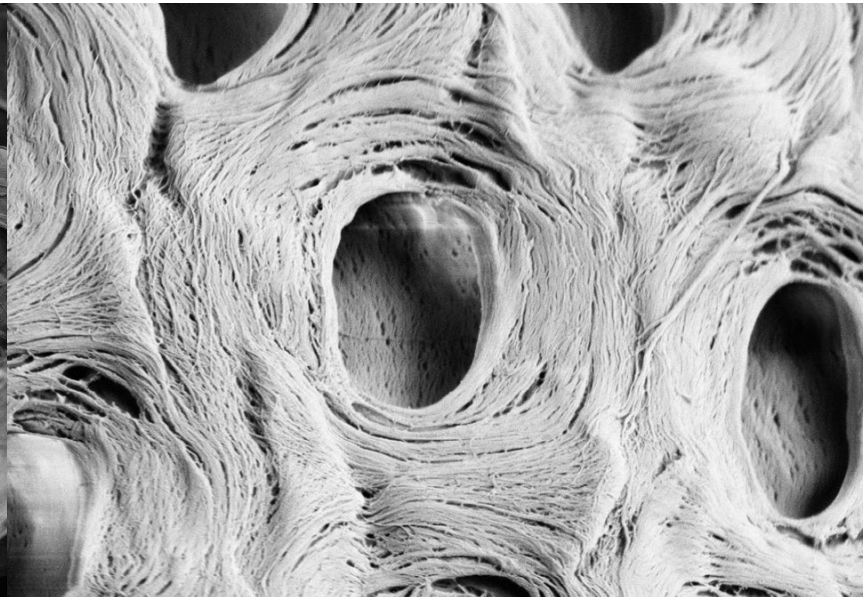
The cell wall is composed of sub-units i.e the hierarchy. Do these swell independently?

Can we “nano-engineer” the cell wall without extracting nanocellulose?



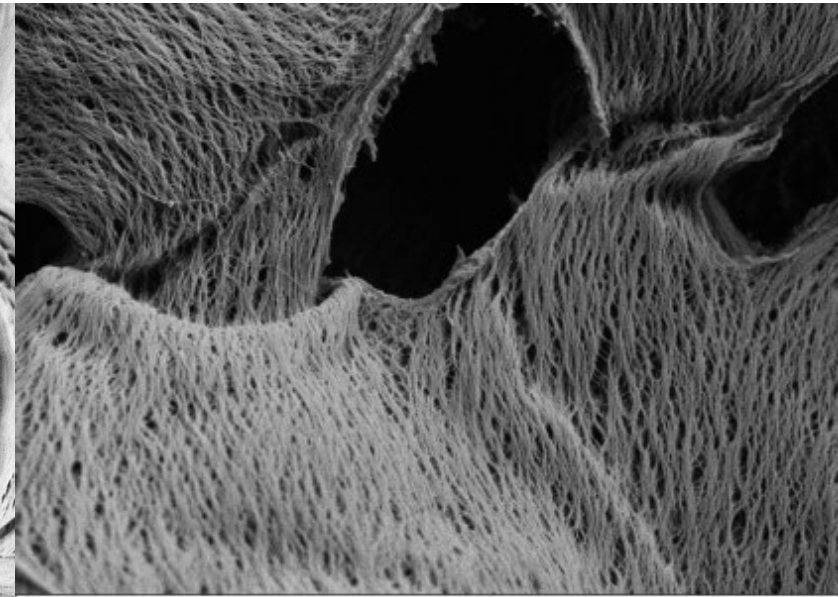
10  $\mu\text{m}$  | EHT = 1.40 kV | Signal A = SE2 | Date : 15 Apr 2015  
WD = 3.4 mm | Mag = 1.53 K X | Time : 18:17:39  
VP Target = 10 Pa

**A!** Aalto University  
Aalto-NMC

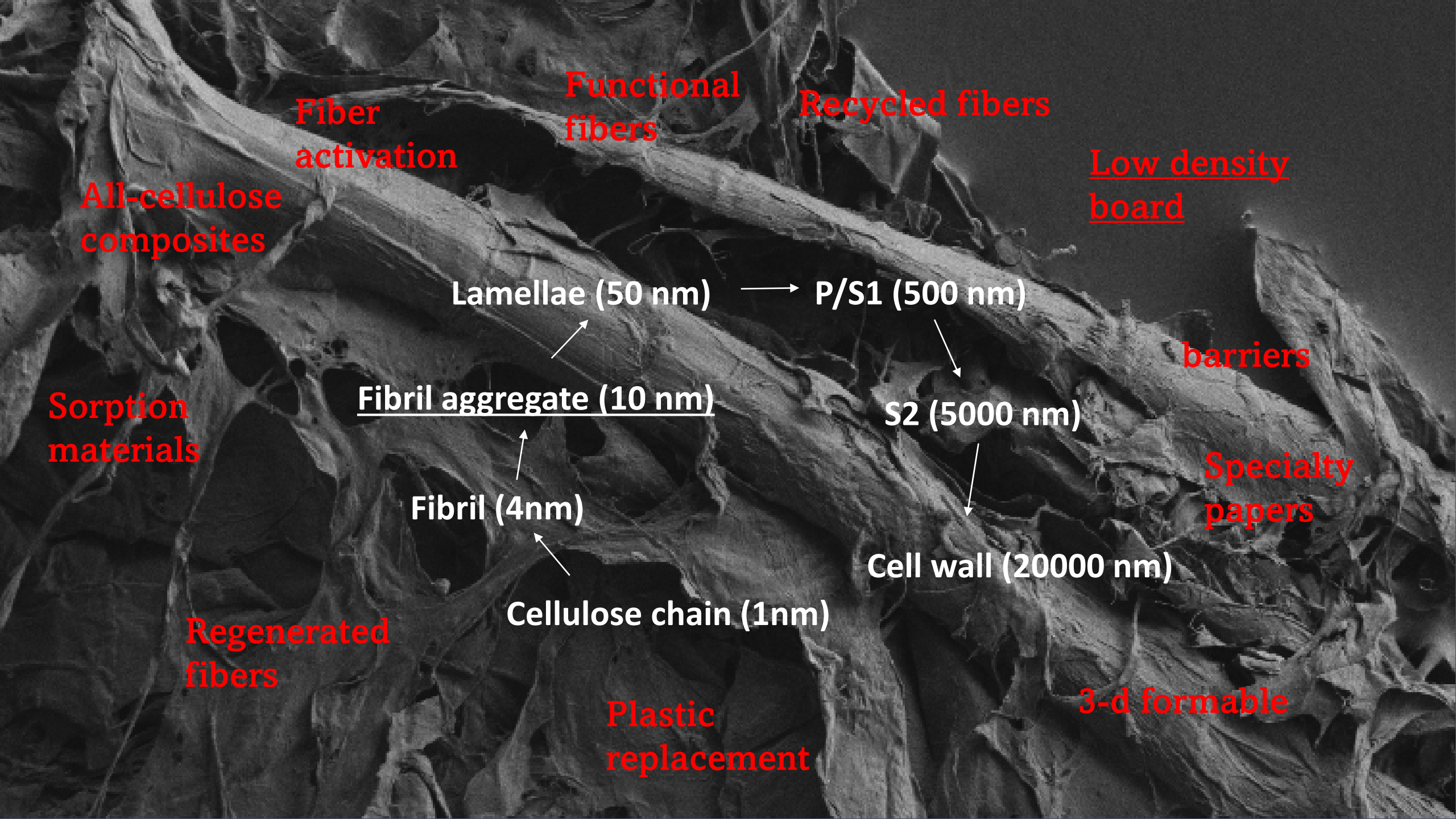


1  $\mu\text{m}$  | EHT = 1.30 kV | Signal A = SE2 | Date : 10 Dec 2015  
WD = 8.0 mm | Mag = 12.27 K X | Time : 15:12:44

**A!** Aalto University  
Aalto-NMC



2  $\mu\text{m}$  | EHT = 1.40 kV | Signal A = SE2 | Date : 15 Apr 2015  
WD = 3.4 mm | Mag = 8.32 K X | Time : 18:19:28  
VP Target = 10 Pa



All-cellulose composites

Fiber activation

Functional fibers

Recycled fibers

Low density board

Sorption materials

Regenerated fibers

Plastic replacement

3-d formable

barriers

Specialty papers

Lamellae (50 nm)

P/S1 (500 nm)

Fibril aggregate (10 nm)

S2 (5000 nm)

Fibril (4nm)

Cell wall (20000 nm)

Cellulose chain (1nm)

# Hypothesis

Macrofibril swelling can be controlled independently from cell wall swelling.

- This can be measured
- Its useful where high specific bond strength is important

# Experimental

## Materials

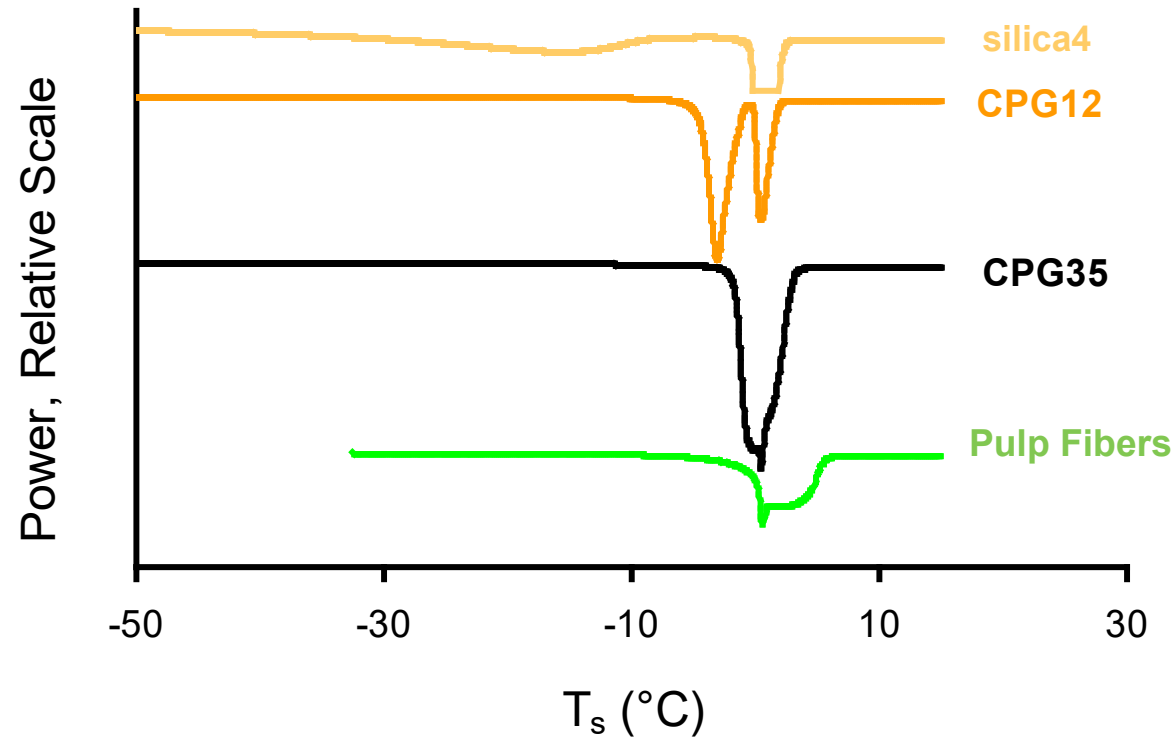
- Pulp: Bleached softwood kraft, never dried, some experiments use an UBSW kraft pulp.
- Functionalization: varying quantities of phosphate groups added to the internal surfaces by drying the pulp from diammonium phosphate/urea aqueous solution under controlled conditions. Exchanged to Na<sup>+</sup> form.

## Analytics

- Water retention value (WRV) for measuring cell wall swelling.
- Se exclusion with 3.2 nm dextran probe for measuring macrofibril swelling.
- Thermoporosimetry for macrofibril swelling, pore size distribution and surface area.
- Molecular dynamics simulation for visualizing macrofibril swelling and estimating surface area (Antti Paajanen).
- X-ray scattering for measure fibril separation (another measure of macrofibril swelling) (Paavo Penttilä, Aleksi Zitting).

In thermoporosimetry, the pore size distribution can be accounted calculated from the well-known Gibbs-Tomson eq., except....

Melting energy of water in for porous glasses and pulp fibers

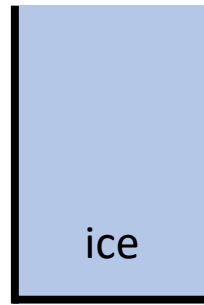
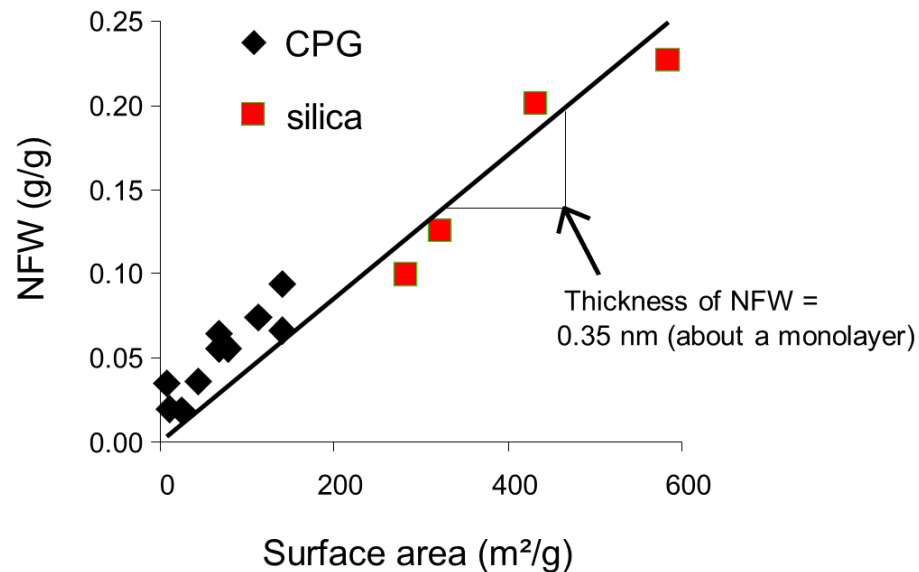


$$D = \frac{k}{\Delta T}$$

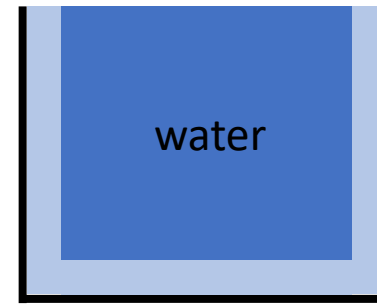
$$V_{\text{pore}} \propto H_m$$

# 2 fractions of nonfreezing water (NFW) in pulp fibers

NFW in mesoporous ( $D > 2\text{nm}$ ) glasses

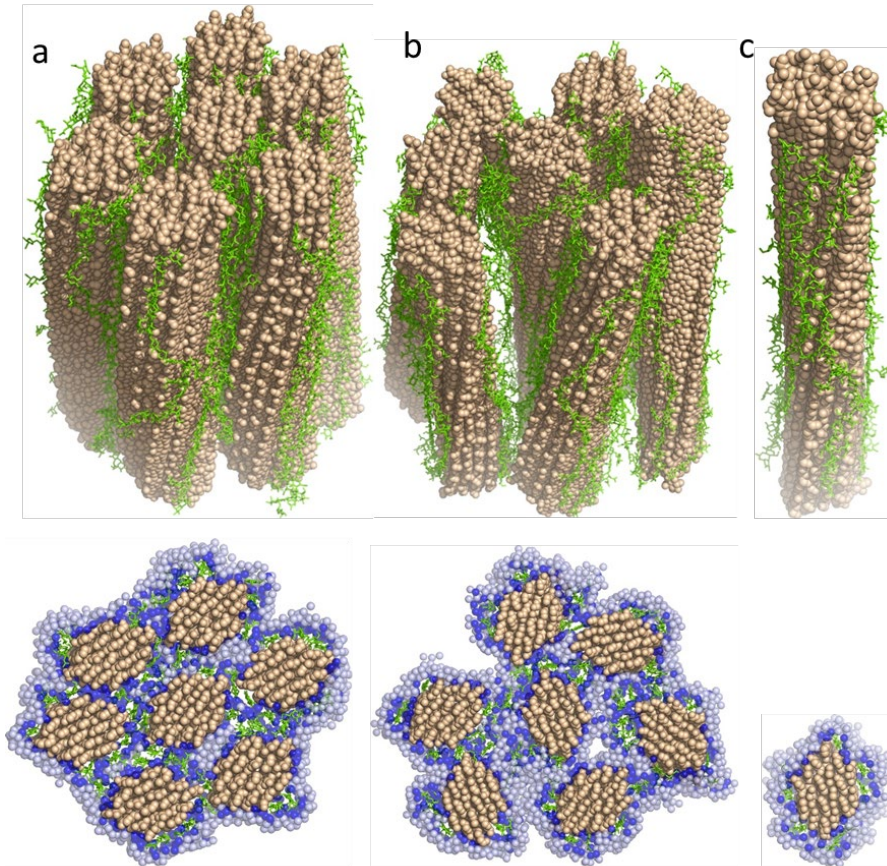


$D < \sim 2\text{ nm}$



$D > \sim 2\text{ nm}$

# Swelling of fibril aggregates



- Molecular dynamics simulation of Spruce S2
- 7 fibril aggregates (macrofibrils with different packing).
- 18 cellulose chains in hexagonal array
- 20% Galactogluccomannan/glucuronoarabinoxylan
- Hornification increases from c to a; deaggregation increases a to c



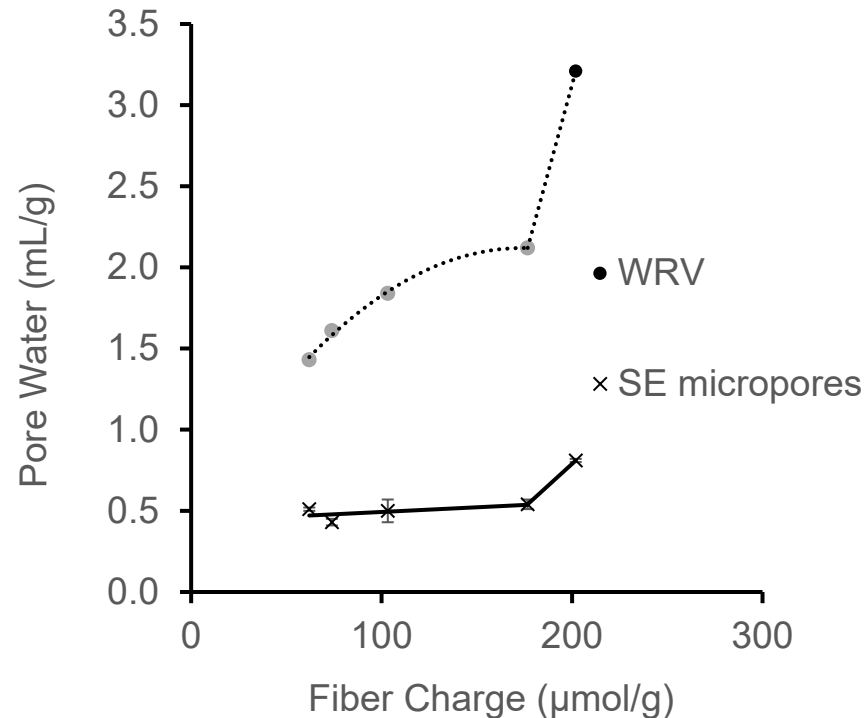
# Surface area and bound water volumes from simulations

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<b>Bond water and surface area</b>	<b>Tight fibril aggregate</b>	<b>Loose fibril aggregate</b>	<b>Single fibril</b>
1 <sup>st</sup> water layer volume, mL/g	0.1585 ± 0.0009	0.197 ± 0.002	0.202 ± 0.004
2 <sup>nd</sup> water layer volume, mL/g	0.205 ± 0.002	0.321 ± 0.007	0.427 ± 0.008
SA, 0.28 nm probe, m <sup>2</sup> /g	1050 ± 6	1490 ± 20	1590 ± 30
SA, 3.2 nm probe m <sup>2</sup> /g*	536 ± 9	624 ± 9	

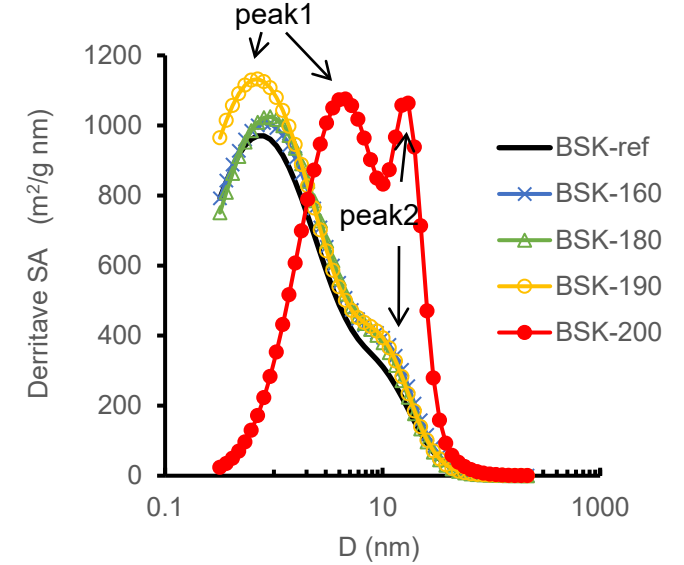
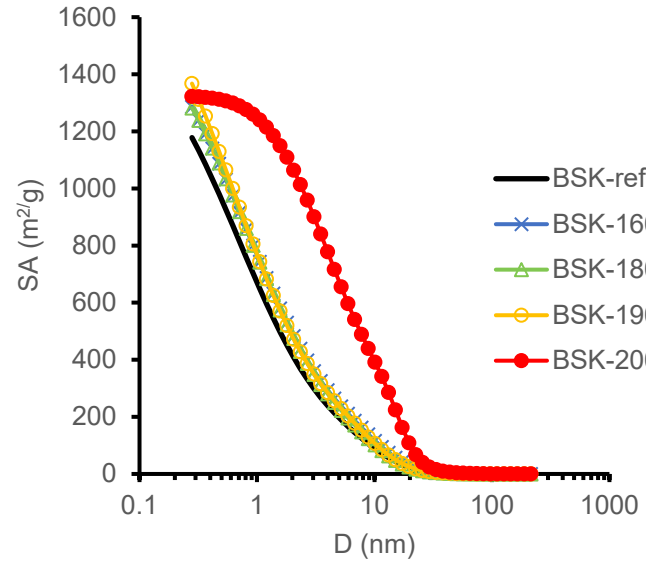
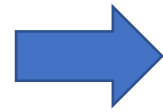
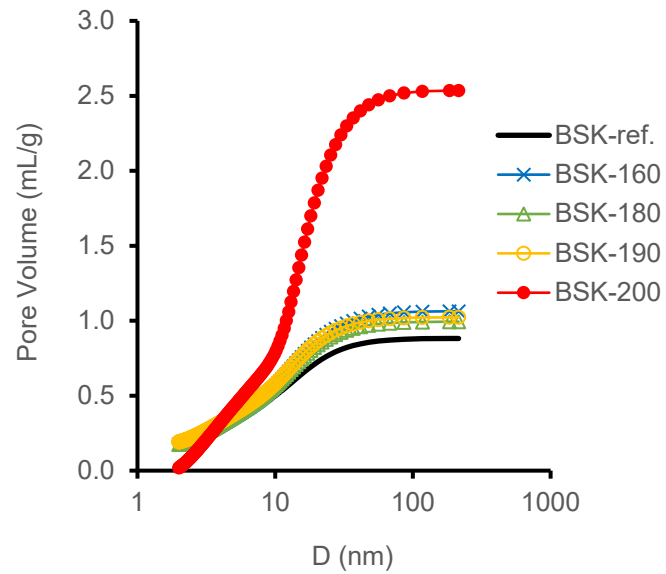
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# Swelling of the cell wall vs swelling of the fibril aggregate

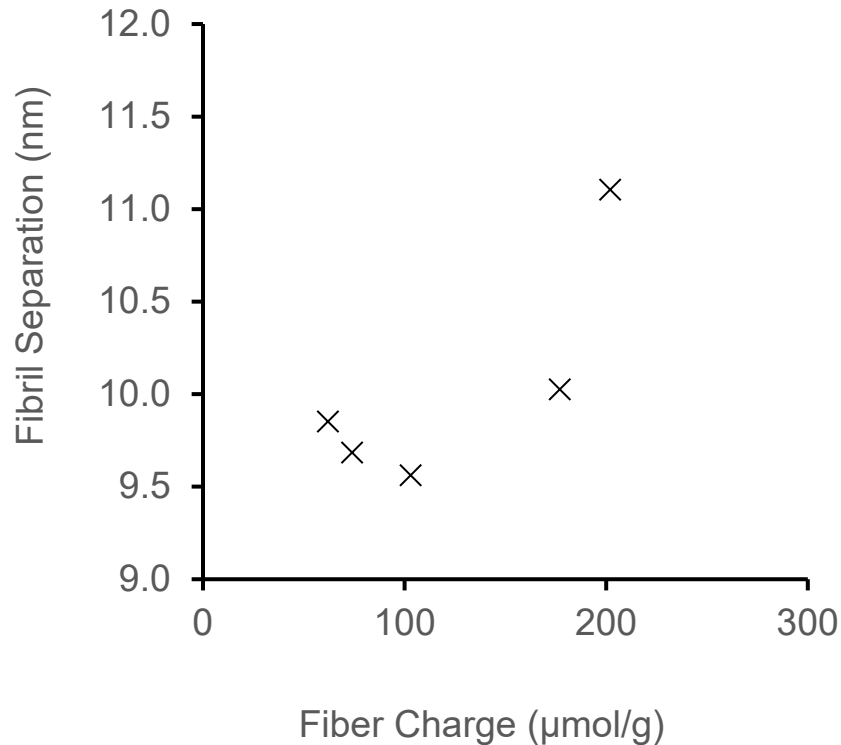


- Adding charge (phosphate groups) increases cell wall swelling elastically up to 180 μmol/g.
- No increase in fibril aggregate swelling in this range.
- At 200 μmol/g charge, the cell wall swells dramatically; fibril aggregate swelling increases 35%.

# The surface area plot shows dramatic effect of macrofibril swelling

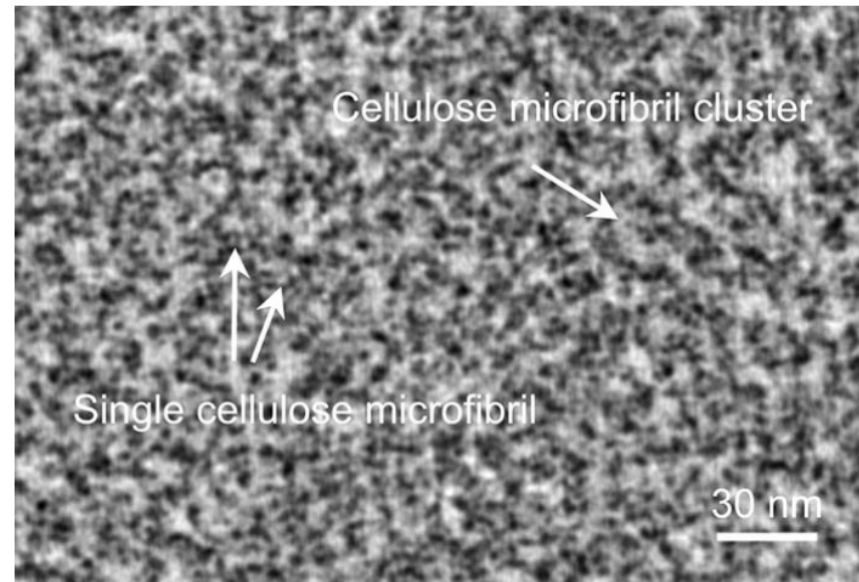


# X-ray scattering of phosphorylated pulps



- Results confirm a distinct increase in fibril separation at the highest charge. i.e. *deaggregation*.
- No presence of cellulose II or other anomalous crystal structures detected.

# Clustering of fibrils in delignified Radiata Pine



**Fig. 1** Tomographic slice image (~ 1 nm thick) of the cross-section of a S2 layer in a delignified early radiata pine wood specimen. The *white/light grey structures* correspond to individual and clustered cellulose microfibrils, and the *surrounding dark grey/black structures* correspond to residual lignin and hemicellulose

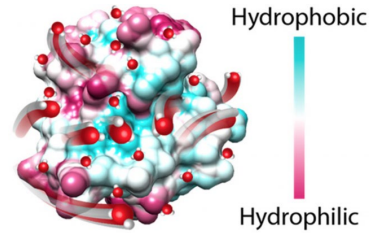
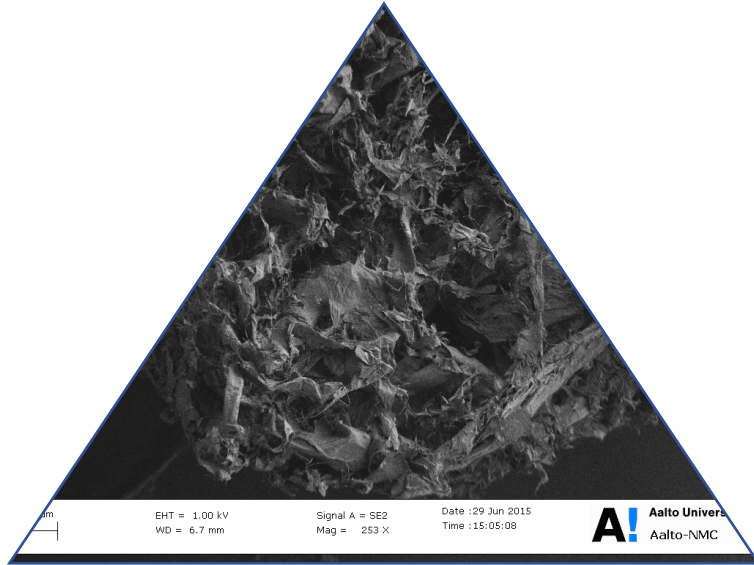
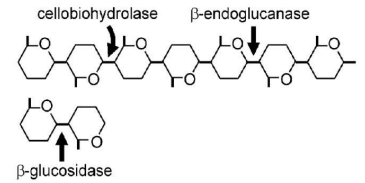
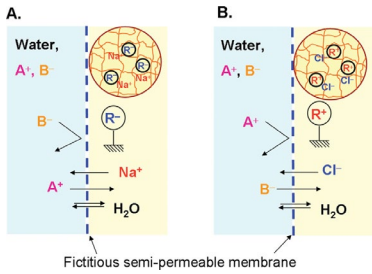
*Xu, P., Donaldson, L. A., Gergely, Z. R. & Staehelin, L. A. Dual-axis electron tomography: A new approach for investigating the spatial organization of wood cellulose microfibrils. Wood Sci. Technol. 41, 101–116 (2007).*

# Hierarchical swelling control allows one to decouple important paper property pairs

Sample	Density	Tensile index	E-Modulus	Light scattering
	kg/m <sup>3</sup>	Nm/g	GPa	(cm <sup>2</sup> /g)
BSK-ref	360	16.6 ± 4.6	3.9	35.5 ± 0.5
BSK-160	519	26.7 ± 3.1	5.7	30.7 ± 1.4
BSK-180	611	43.0 ± 2.4	3.8	24.9 ± 0.6
BSK-190	611	44.9 ± 7.9	13.7	24.8 ± 0.3
BSK-200	614	59.7 ± 11	16.7	19.1 ± 4.9

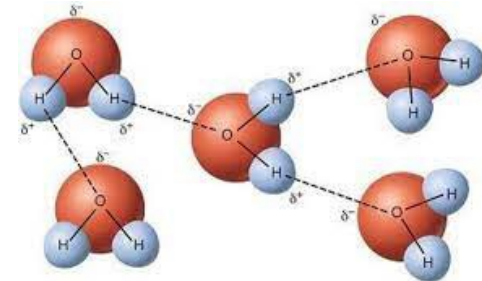
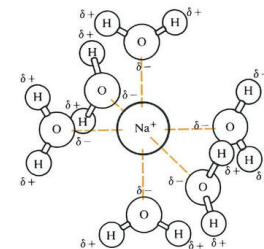
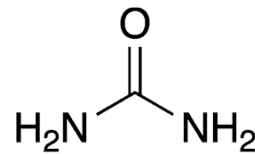
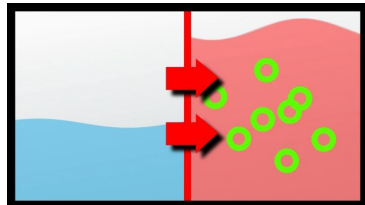
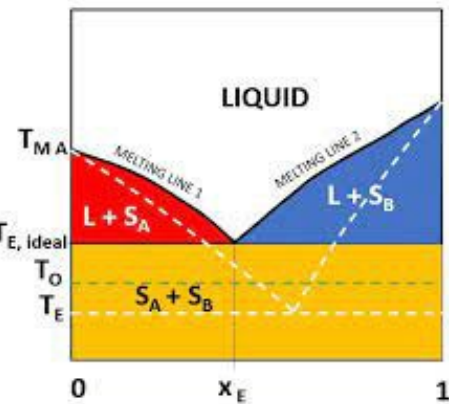


# Controlling the hierarchical swelling

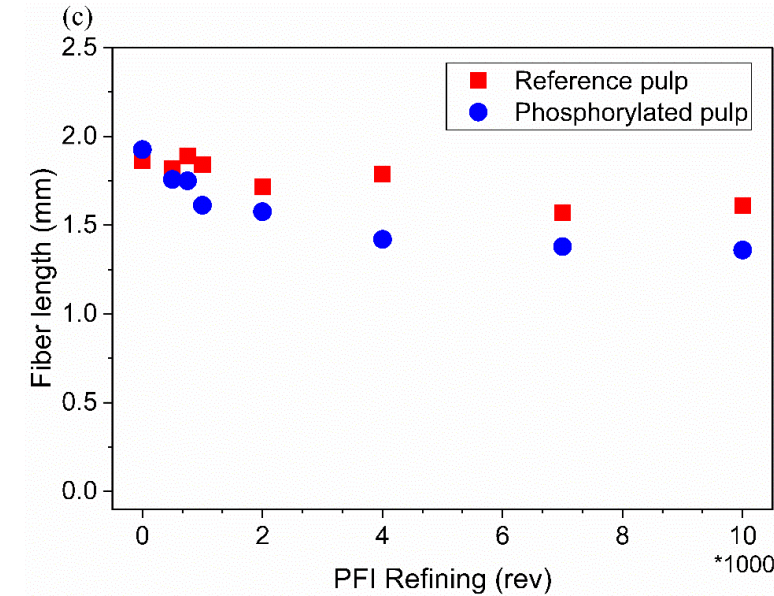
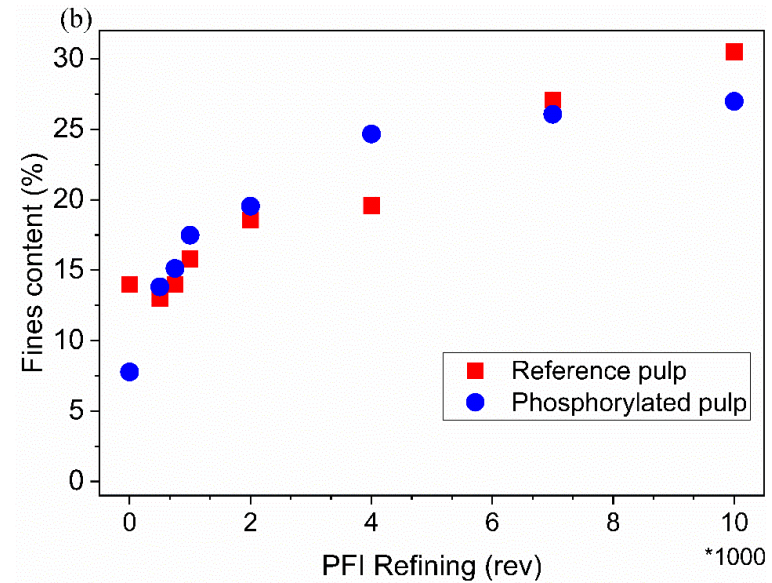
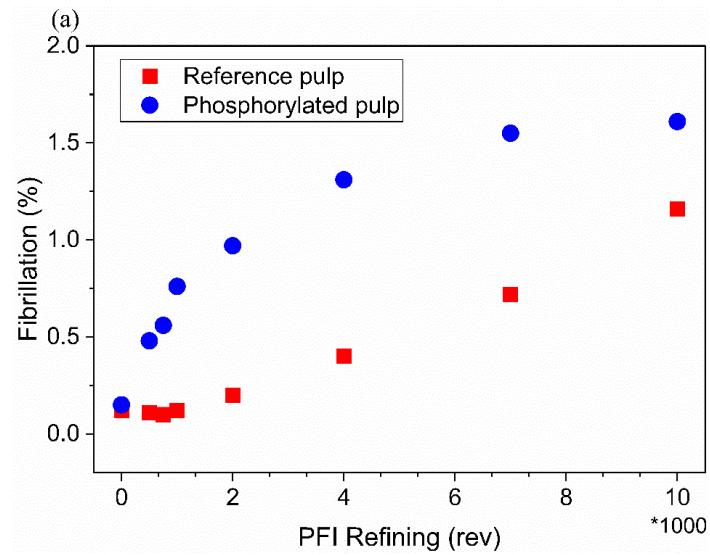


H-bond disruptors

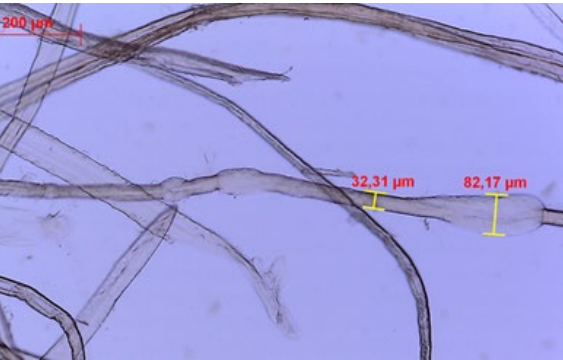
Osmotic pressure



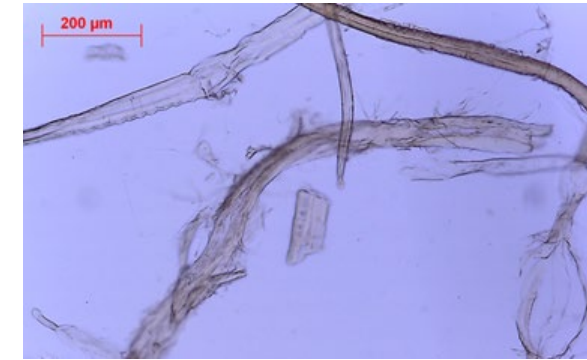
# Phosphorylated pulps fibrillate but don't degrade in refining



*Phosphorylated USWK, unrefined*



*Phosphorylated USWK, refined*

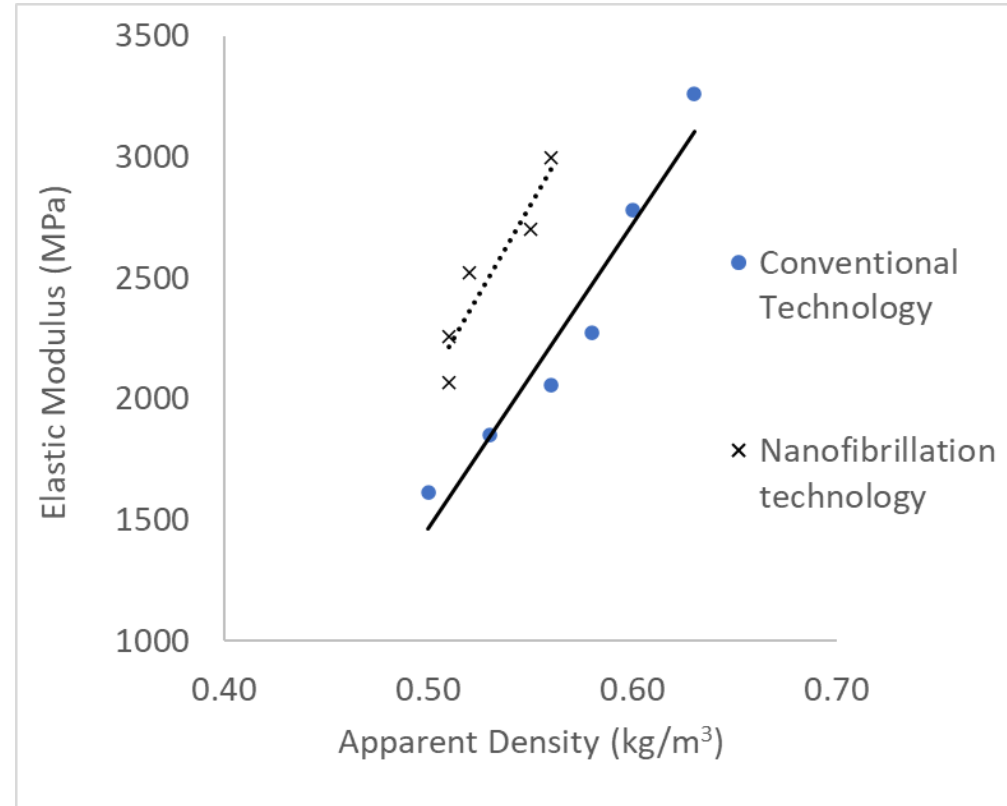




# High strength/density paper by controlling hierarchical swelling



- SA or macrofibril  $\sim 500$  m<sup>2</sup>/g
- SA microfibril  $\sim 1600$



# Conclusions

- Its possible to control the macrofibril swelling independently from the cell wall swelling and more broadly to manipulate the swelling hierarchy of pulp fibers.
- Increased electrostatic repulsion, disruption Van der Waals and H-bonds, mechanical shear are relevant tools.
- Thermoporosimetry is a good tool for measuring the fiber structural effects.
- In some cases, nano-engineering the fiber is more favorable than producing and adding nanocellulose.

# Allocation of nonfreezing water between micropores and mesopores

Sample	Total NFW (mL/g)	Monolayer NFW in mesopores (mL/g)	NFW in subfreezing micropores (mL/g)
BSK-ref	$0.25 \pm 0.07$	0.10	0.15
BSK-160	$0.27 \pm 0.06$	0.11	0.16
BSK-180	$0.27 \pm 0.05$	0.12	0.16
BSK-190	$0.29 \pm 0.08$	0.11	0.18
BSK-200	$0.27 \pm 0.03$	0.27	0.00